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Effects of extrusion process on *Fusarium* and *Alternaria* mycotoxins in whole grain triticale flour

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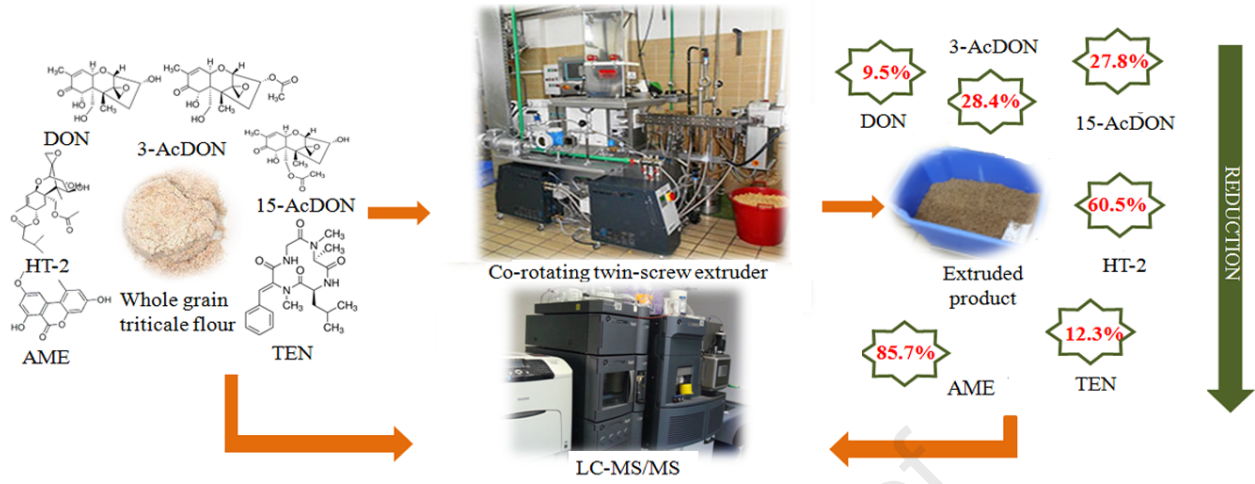
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1 **Effects of extrusion process on *Fusarium* and *Alternaria* mycotoxins in whole grain**  
2 **triticale flour**

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## 11 **Abstract**

12 Effects of extrusion processing parameters of co-rotating twin-screw extruder – screw speed  
13 (*SS*= 500, 650, 800 rpm), feed rate (*FR*= 22, 26, 30 kg/h), and moisture content of the material  
14 (*MC*= 20, 25, 30 g/100 g), on the reduction rate of deoxynivalenol (DON), 3- and 15-  
15 acetyldeoxynivalenol (3- and 15-AcDON), HT-2 toxin (HT-2), tentoxin (TEN) and alternariol  
16 monomethyl ether (AME), in whole grain triticale flour were investigated, together with the  
17 physico-chemical characterization of obtained products. The die temperature of the extruder  
18 ranged between 113 and 151 °C, the pressure at the die was from 2.7 to 7.9 MPa, the mean  
19 retention time of material in the barrel was between 4 and 11 s, torque ranged between 39.6 to  
20 59.4 Nm, while the specific mechanical energy ranged from 66.9 to 125 kWh/t. Optimal  
21 parameters for lowering the concentration of each investigated mycotoxins were: *SS*= 650  
22 rpm, *FR*= 30 kg/h, *MC*= 20 g/100 g, with a reduction of 9.5, 27.8, 28.4, 60.5, 12.3 and 85.7%  
23 for DON, 3-AcDON, 15-AcDON, HT-2, TEN and AME, respectively. Present study is the  
24 first report for the fate of mycotoxins (3-AcDON, 15-AcDON, HT-2, TEN and AME) studied  
25 less during extrusion process of naturally contaminated whole grain triticale flour.

26  
27 **Keywords:** whole grain triticale flour, co-rotating twin-screw extruder, mycotoxins reduction,  
28 LC-MS/MS.

## 33 **Abbreviations**

34 *DON* – deoxynivalenol

35 *3-AcDON* – 3-deoxynivalenol

- 36 *15-AcDON* – 15-deoxynivalenol
- 37 *HT-2* – HT-2 toxin
- 38 *TEN* – tentoxin
- 39 *AME* – alternariol monomethyl ether
- 40 *BD* – bulk density (g/mL)
- 41 *FR* – feed rate (kg/h)
- 42 *MC* – moisture content (g/ 100 g)
- 43 *P* - pressure at the die (MPa)
- 44 *PH* – pellet hardness (kg)
- 45 *RT* – retention time in the barrel (s)
- 46 *SME* – torque (Wh/kg)
- 47 *SS* – screw speed (rpm)
- 48 *Tend* – die temperature (°C)
- 49 *WAI*- water absorption index (g/g)
- 50 *WSI* – water solubility index (g/100 g)
- 51 *rAME* – reduction of alternariol monomethyl ether (%)
- 52 *rDON* – reduction of deoxynivalenol (%)
- 53 *r15-AcDON* – reduction of 15-acetyldeoxynivalenol (%)
- 54 *r3-AcDON* – reduction of 3-acetyldeoxynivalenol (%)
- 55 *rHT-2* – reduction of HT-2 toxin (%)
- 56 *rTEN* – reduction of tentoxin (%)

57

## 58 **1. Introduction**

59 Cereals worldwide are at risk to be contaminated by mycotoxins both in the field as the  
60 result of infection by different fungi or after harvest, as a consequence of ineffective drying or

61 poor storage conditions. Mycotoxins are toxic secondary metabolites of filamentous fungi,  
62 mainly *Aspergillus* species (spp.), *Penicillium* spp. and *Fusarium* spp. (Agriopoulou,  
63 Stamatelopoulou, & Varzakas, 2020; Oliveira, Zannini, & Arendt, 2014). Among all of the  
64 fungal secondary metabolites currently known, only a few groups of mycotoxins are  
65 important from the safety and economic points of view; namely aflatoxins (AFs), mainly  
66 produced by *Aspergillus* spp., ochratoxin A (OTA), produced by *Aspergillus* and *Penicillium*  
67 spp., and zearalenone (ZEN), fumonisins (FBs) and trichothecenes (deoxynivalenol (DON),  
68 T-2 and HT-2 toxin (T-2, HT-2), diacetoxyscirpenol (DAS)), primarily produced by many  
69 *Fusarium* spp. (Agriopoulou et.al., 2020; Streit et al., 2012). In recent years, less studied  
70 *Alternaria* toxins gained more and more interest due to their possibility to cause toxic effects  
71 on animal and human health (EFSA, 2016). *Alternaria* spp. produces around 70 different  
72 mycotoxins, but the most relevant are tenuazonic acid (TeA), tentoxin (TEN), alternariol  
73 (AOH), alternariol monomethyl ether (AME), and altenuene (ALT). Consumers are mainly  
74 exposed to *Alternaria* toxins through processed foods or fruits (EFSA, 2016). The presence of  
75 mycotoxins in cereals is recognized as a worldwide concern since cereals represent one of the  
76 main parts of the human diet and animal nutrition (Babič et al., 2021; Janić Hajnal et al.,  
77 2019;). Since, mycotoxins are heat stable, during common processing methods of cereals  
78 (primary and secondary processes), reduction of their content may have occurred, but they  
79 may not be destroyed (Bullerman, & Bianchini, 2007; Agriopoulou et.al., 2020; Wan,  
80 Bingcan, & Rao, 2020 ). Postharvest approaches for the reduction of mycotoxins are  
81 important topics in food safety research. Various methods (physical, chemical, and biological)  
82 have been applied to prevent mycotoxin production, or reduce mycotoxin content (Liu et al.,  
83 2020). According to previously published studies recently reviewed by Schaarschmidt &  
84 Fahl-Hassek (2018, 2021), extrusion could be effective as a physical detoxification approach  
85 in reducing some mycotoxins in wheat and maize (Liu et al., 2020; Schaarschmidt & Fahl-

86 Hassek, 2018; 2021; Wan et al., 2020). The potential reduction of aflatoxin B1, B2, G1, and  
87 G2 levels by extrusion in corn-based products was investigated by Massarolo et al. (2021).  
88 The extrusion process combines a high temperature, high pressure, and short time process and  
89 can be used for the production of a range of cereal products and animal feeds. Generally, the  
90 extrusion process results in chemical changes and modifications (protein denaturation, starch  
91 gelatinization, polymer cross-linking, Maillard reactions, etc.), both for food components and  
92 present contaminants (Singha, Singh, Muthukumarappan, & Krishnan 2018; Torbica, Belović,  
93 Popović, & Čakarević, 2021). However, the extent of mycotoxin contamination reduction in a  
94 finished product depends on several factors, including the type of extruder, the extrusion  
95 conditions (extruder temperature, screw speed, feed rate, pressure, and residence time in the  
96 extruder), moisture content of the raw materials or extrusion mixture, chemical structure of  
97 mycotoxins, its initial content in the raw material, as well as depend on the potential matrix  
98 effects (Schaarschmidt & Fauhl-Hassek, 2018, 2021; Wan et al., 2020 ). The stability of  
99 mycotoxins during extrusion and the ability of extrusion processes to reduce the content of  
100 mycotoxins in extruded products have been studied to promote the degradation of the  
101 mycotoxins, mostly *Fusarium* toxins (Schaarschmidt & Fauhl-Hassek, 2018; 2021). The first  
102 report presenting the possibility of reduction of *Alternaria* toxins in wheat by extrusion was  
103 published by Janić Hajnal et al.(2016). The studies published so far regarding the possibilities  
104 of mycotoxins reduction during the extrusion process mostly referred to maize and wheat  
105 (Schaarschmidt & Fauhl-Hassek, 2018; 2021), rarely to barley, oats, and rice, while there is  
106 almost no data available related to the possibility of reducing of mycotoxins in triticale grain  
107 (*×Triticosecale*) during processing. One of the possible reasons is that triticale (produced by  
108 cross-breeding wheat and rye) is a less represented cereal in the world, and another reason is  
109 that it has been mostly used as animal feed, and less for human nutrition and biofuel  
110 production (Gagiu, 2018). However, the world production of triticale has kept increasing



111 during the last few years. Between 2000 and 2019, the largest cultivating countries of triticale  
112 grain were: Poland, Germany, France, Belarus, China ( $\leq 201,870,707$  t in total), Hungary,  
113 Australia, Lithuania, Russian Federation, Spain ( $\leq 32,169,927$  t in total); Austria, Czechia,  
114 Sweden, Romania, Denmark ( $\leq 18,168,817$  t in total) and Turkey, Chile, Brazil, Serbia,  
115 Switzerland ( $\leq 7,755,711$  t in total). The Republic of Serbia is among the 20 largest producers  
116 of triticale in the world, with an annual production of 102,231 tons in 2019 (FAO, 2019).  
117 Considering that triticale is often contaminated with mycotoxins (Gagiu, 2018), in this study  
118 the influence of extrusion parameters on reduction of examined mycotoxins content in whole  
119 grain triticale flour was investigated.

120 Modern mathematical approaches such as Response Surface Methodology (RSM) to optimize  
121 the extrusion process (Singha & Muthukumarappan, 2017; Kojić et. al., 2019) can be used to  
122 regulate the quality of the extrudate and evaluate the effect of extrusion variables on the  
123 reduction of mycotoxins. The present work aims to optimize the extrusion process and  
124 evaluate the effect of different extrusion process variables (screw speed, feed rate, and  
125 moisture content of the material) on the quality of extrudates and on the reduction of  
126 mycotoxins using whole grain triticale flour that was naturally contaminated with mycotoxins.  
127 *Fusarium* and also less studied *Alternaria* toxins were investigated in the study. This work  
128 provides for the first time important data on the reduction effect of *Fusarium* and *Alternaria*  
129 toxins during the extrusion process, which is commonly used for the production of animal  
130 feed and food.

## 131 **2. Material and methods**

### 132 *2.1 Material*

133 Approximately 300 kg of triticale grain ( $\times$ *Triticosecale*) naturally contaminated by  
134 mycotoxins was provided by the Institute of Field and Vegetable Crops, Novi Sad (Serbia)  
135 and finely ground using a hammer mill (model 9FQ-50, XT Machinery, China) driven by 22

136 kW electric motor and equipped with 16 hammers arranged in four rows and with the sieve of  
137 1 mm diameter. To achieve an adequate homogeneity level of the ground material before  
138 sampling for the analysis and the extrusion processing, the whole grain triticale flour was  
139 mixed in a Muyang SLHSJ0.2A double-shaft paddle mixer (Muyang, Yangzhou, China) for  
140 90 s. Mixing homogeneity of triticale flour was assured by Microtracer® method, using  
141 external tracers for mixing homogeneity testing (Clark, Behnke, & Poole, 2007), and also,  
142 twelve subsamples were taken for investigations of different mycotoxins levels in whole grain  
143 triticale flour.

144

## 145 *2.2 Extrusion conditions*

146 Co-rotating twin-screw extruder (Bühler BTSK-30, Bühler, Uzwil, Switzerland) with a  
147 total barrel length of 880 mm consisted of 7 sections and length/diameter ratio of 28:1 was  
148 used for the extrusion of the ground triticale grain. The extruder was equipped with two  
149 tempering tools for controlling water temperature for jacketed heating/cooling of barrel's  
150 sections. The first tempering tool controlled the temperature of sections 2, 3, and 4 (60 °C)  
151 while another tempering tool was used for controlling the temperature of sections 6 and 7 (set  
152 at 100 °C). The die plate with one 6 mm diameter opening and cone inlet (total die open area  
153 of 28.26 mm<sup>2</sup>) was used. In this experiment, the same screw configuration was used as  
154 presented by Kojić et al. (2019). The scheme of the co-rotating twin-screw extruder is  
155 presented in Fig. 1. Screw speed, feed rate, and moisture content of the material in the  
156 extruder barrel were varied during extrusion according to the applied experimental design  
157 (Table 1). A total of fifteen extruded samples were obtained (TS-1 to TS-15). Targeted  
158 moisture content in the barrel was achieved by adding water at the end of section 1 of the  
159 barrel using a cavity pump. Sensors for measuring the pressure and temperature of the die  
160 were positioned at the die head. All extrusion data, including die temperature, pressure at the

161 die, motor load, and specific mechanical energy were read directly from the PLC screen of the  
162 extruder. The final length of the product was obtained by the rotational knife that faced the die  
163 outlet and was fitted with six knives, with a rotational speed set at 1100 rpm. Drying and  
164 subsequent cooling of the extrudates were done in a fluidized bed vibro dryer/cooler (model  
165 FB 500 x 2000, Amandus Kahl GmbH & Co. KG, Germany).

166

### 167 2.3 *Chemicals and reagents*

168 A mixed trichothecene standard solution in acetonitrile (DON, 3-AcDON, 15-AcDON, T-2,  
169 HT-2, DAS), produced by Trilogy (Washington, MO, USA) and individual standards of TEN,  
170 AOH, AME, ZEN, OTA, FB1, and FB2 (Romer Labs, Tulln, Austria) were used. The stock  
171 standard solutions and the working standard solutions were prepared in acetonitrile and stored  
172 in amber glass vials at  $-20\text{ }^{\circ}\text{C}$ . The certified purity of individual standard substances was  
173 between  $98.5 \pm 1.5\%$  and  $99.5 \pm 0.5\%$ . Working standard solutions of known concentrations  
174 were prepared by the appropriate dilution of the stock standard solution. Acetonitrile,  
175 methanol (Honeywell, Seelze, Germany), acetic acid (Sigma-Aldrich, Steinheim, Germany),  
176 and ammonium acetate (Merck, Darmstadt, Germany) were of pro analysis or LC-MS purity.  
177 Deionized water was prepared with a Milli-Q system (Millipore, Bedford, MA, USA).

178

### 179 2.4 *Moisture content*

180 Moisture content in whole grain triticale flour sample and extruded product samples was  
181 determined according to ISO method (ISO, 2009), and expressed on a dry weight basis.

182

### 183 2.5 *Bulk density of extrudates*

184 The bulk density (BD) of each extruded product sample was measured with a bulk density  
185 tester (Tonindustrie, West und Goslar, Germany) in triplicate.

186

187 *2.6 Hardness determination*

188 The hardness of the extrudates (PH) was determined using a Texture Analyser (model  
189 TA.HDPlus, Stable Micro Systems Ltd, Godalming, Surrey, UK) equipped with the 50 kg  
190 load cell. The single extrudate was diametrically positioned between a plate and movable  
191 cylindrical probe (diameter 45 mm). Test settings were as follows: pretest speed: 2.0 mm/s;  
192 test speed: 0.16 mm/s; post-test speed: 10 mm/s; distance: 2.5 mm; trigger force: 100 g. The  
193 maximum peak force from the force-time graph was considered as an indication of hardness.  
194 Hardness was expressed in kg as the mean of the results of 20 extrudates from each trial.

195

196 *2.7 Water Absorption Index and Water Solubility Index*

197 Water absorption index (WAI) and water solubility index (WSI) were determined by the  
198 method of Anderson, Conway, & Peplinski (1970) with slight modification. In brief, 0.2 g of  
199 ground extrudates was suspended in 5 mL of distilled water in weighed 15 mL glass  
200 centrifuge tube. The tube was stirred on a Vortex mixer (VELP Scientifica Srl, Italy) for 2  
201 min and then centrifuged (Eppendorf Centrifuge 5804 R, Hamburg, Germany) at 5000 × g for  
202 20 min at room temperature (25 °C). The supernatant was decanted into an evaporating dish  
203 of known weight. The gel obtained after decantation of the supernatant was measured and  
204 WAI was calculated using equation (1):

$$205 \quad WAI \text{ (g/g)} = \textit{weight of gel} / \textit{weight of sample} \quad (1)$$

206 The WSI was determined using equation (2) from the weight of dry solids after evaporation of  
207 supernatant from the WAI test at 105 °C in drying oven (UNB 400, Memmert, Germany):

$$208 \quad WSI \text{ (g/100g)} = \textit{weight of dissolved solids in the supernatant} / \textit{weight of sample} \times 100 \quad (2)$$

209 WAI and WSI were expressed as the mean of the results of four repetitions from each trial.

210

211 *2.8 Sample preparation for LC-MS/MS analysis*

212 The sample preparation procedure consisted of a simple one-step sample extraction which  
213 was previously described in detail by Babič et al. (2021), Topi, Tavčar-Kalcher, Pavšič-Vrtač,  
214 Babič, & Jakovac-Strajn (2019) and Topi, Babič, Pavšič-Vrtač, Tavčar-Kalcher, & Jakovac-  
215 Strajn (2021).

216

217 *2.9 LC-MS/MS analysis*

218 For the determination of 13 mycotoxins (DON, 3-AcDON, 15-AcDON, DAS, HT-2, T-2,  
219 ZEN, FB1 and FB2, OTA, AOH, AME, and TEN) ultra-performance liquid chromatography  
220 coupled with a triple-quadrupole mass spectrometer (UPLC-MS/MS) was used with  
221 electrospray ionization (ESI) interface and MassLynx software for data collection and  
222 processing (Waters, Milford, MA, USA). The LC-MS/MS method for quantification of above  
223 mentioned mycotoxins was described, in detail by Babič et al. (2021), Hojnik et al.(2019),  
224 Topi et al. (2019; 2021).

225

226 *2.10 Method validation*

227 For detected mycotoxins in whole grain triticale flour (DON, 3-AcDON, 15-AcDON, HT-  
228 2, TEN, and AME), the method was validated in terms of matrix effect, linearity, trueness,  
229 precision, limit of detection (LOD), and limit of quantification (LOQ) by an in-house quality  
230 control procedure, in the manner described in detail in our previous studies (Janić Hajnal et  
231 al., 2015; 2016; 2019), for both matrices (whole grain triticale flour and extruded product  
232 samples). The used spiking levels (four) of each mycotoxin (DON, 3-AcDON, 15-AcDON,  
233 HT-2, TEN, and AME) into both matrices (whole grain triticale flour and extruded product  
234 samples) for method validation are presented in Table 2 and Table 3. The LOD of the single

235 analytes was determined at a signal-to-noise ratio of 3:1. A value 3.3 times the LOD was  
236 selected as the LOQ.

237

## 238 *2.11 Statistical analysis*

### 239 *2.11.1 Principal component analysis (PCA)*

240 Principal component analysis (PCA) was conducted to elucidate and identify the acquired  
241 data. The analysis of variance (ANOVA) was accomplished, with a particular purpose to  
242 inquire the effects of the factor variables over the responses. The calculation of the ANOVA,  
243 based on the gathered experimental results was done using StatSoftStatistica 13.3® software  
244 (Statistica, 2013).

245

### 246 *2.11.2 Response surface methodology (RSM)*

247 The impact of the three extrusion factor variables: *SS* (500, 650, and 800 rpm), *FR* (22, 26,  
248 and 30 kg/h), and *MC* (20, 25, and 30 g/100 g) on the extrusion of the whole grain triticale flour  
249 were investigated according to the experimental plan presented in Table 1. The scopes of these  
250 factors were ascertained by the preliminary trial. The experimental data employed for the  
251 analysis were produced by the Box and Behnken (BB) experimental design, which was utilized  
252 to restrict the sample size to 15 which was adequate to assessing second order polynomial  
253 (SOP) coefficients (Singha & Muthukumarappan, 2017).

254

### 255 *2.11.3 Standard score*

256 Standard scores were evaluated for different mycotoxins reduction trials, according to the  
257 applied extruding process. The ranking method was based according to the ratio of the raw  
258 data and the extreme values for each response (Kojić et al., 2019), following the equation (3):

$$\bar{x}_i = \frac{x_i - \min_i x_i}{\max_i x_i - \min_i x_i}, \forall i, \text{ where } x_i \text{ represents the raw data.} \quad (3)$$

### 260 3. Results and discussion

#### 261 3.1 Evaluation of the LC-MS/MS method

262 The validation data of the analytical method for the determination of quantified  
 263 mycotoxins are given in Table 2. All investigated mycotoxins showed slight signal  
 264 suppression, except for AME, which showed strong signal suppression in both matrices  
 265 (whole grain triticale flour and extruded product samples). The other exception relates to the  
 266 HT-2 toxin, which showed slight signal enhancement in extrusion product samples. Method  
 267 exhibited good linearity, with linear regression coefficient ( $r^2$ ) above 0.9945.

268 Through recovery studies, the trueness of the analytical method was evaluated. The  
 269 apparent recoveries ( $R_A$ ) and the sample preparation recoveries ( $R_E$ ) for target analytes were  
 270 calculated as described in our previous studies (Janić Hajnal et al. 2015; 2016; 2019). It can  
 271 be seen (Table 2) that the  $R_A$  and the  $R_E$  for all target analytes were above 70% in both  
 272 matrices. The only exception showed AME for  $R_A$ , for the reason that AME strongly  
 273 suppresses the analytical signal.

274 Repeatability and within-laboratory reproducibility were used for expression of the  
 275 precision of method used, for both whole grain triticale flour and extruded product samples.  
 276 Precision gave relative standard deviation (RSD) values within the range of 1.9 – 14.6% and  
 277 2.6 – 18.0%, respectively, fulfilling the criteria of  $RSD \leq 20\%$  and indicating a good precision  
 278 of the method used (Table 3).

279 LODs and LOQs for both matrices (whole grain triticale flour and extruded product  
 280 samples) were as follows: 15  $\mu\text{g/kg}$  and 50  $\mu\text{g/kg}$  for DON, 0.9  $\mu\text{g/kg}$  and 3  $\mu\text{g/kg}$  for 3-  
 281 AcDON, 15-AcDON and HT-2 toxin, and 3.8  $\mu\text{g/kg}$  and 12.5  $\mu\text{g/kg}$  for TEN and AME,  
 282 respectively.

283

284 *3.2 Determination of mycotoxins content*

285 The examined mycotoxins were quantified by an external matrix-matched calibration  
286 procedure (separate calibrations were prepared for both whole grain triticale flour and  
287 extruded product samples), to compensate for the matrix effects. The following mycotoxins  
288 were quantified: DON, 3-AcDON, 15-AcDON, HT-2, TEN, and AME. The results obtained  
289 were corrected for sample preparation recovery ( $R_E$ ) and were expressed on a dry matter  
290 basis. Initial water content on a dry weight basis was 10.9 g/100 g in naturally contaminated  
291 whole grain triticale flour, while initial concentrations (average values of twelve  
292 measurements) of quantified mycotoxins expressed on a dry matter basis were  $274.4 \pm 36.4$   
293  $\mu\text{g/kg}$ ,  $2.86 \pm 0.24 \mu\text{g/kg}$ ,  $4.86 \pm 0.43 \mu\text{g/kg}$ ,  $4.59 \pm 0.42 \mu\text{g/kg}$ ,  $29.8 \pm 1.78 \mu\text{g/kg}$ , and  $16.7 \pm$   
294  $5.37 \mu\text{g/kg}$ , for DON, 3-AcDON, 15-AcDON, HT-2, TEN and AME, respectively. All  
295 extruded product samples were analyzed in duplicate.

296 The water content of extruded product samples was ranged from 8.05 to 14.1 g/100 g on a  
297 dry weight basis, while the final concentration expressed on a dry matter basis of quantified  
298 mycotoxins in extruded product samples were ranged from 229.0 to 274.1  $\mu\text{g/kg}$  for DON,  
299 from 1.12 to 2.81  $\mu\text{g/kg}$  for 3-AcDON, from 2.60 to 4.47  $\mu\text{g/kg}$  for 15-AcDON, from 1.81 to  
300 3.47  $\mu\text{g/kg}$  for HT-2, from 23.5 to 29.3  $\mu\text{g/kg}$  for TEN and from 1.37 to 7.83  $\mu\text{g/kg}$  for AME.

301

302 *3.3 Reduction of mycotoxins by extrusion processing*

303 The effects of extrusion process variables – screw speed (SS), feed rate (FR), and moisture  
304 content (MC) – on observed responses (DON, 3-AcDON, 15-AcDON, HT-2, AME, and TEN  
305 reduction rate,  $P$ ,  $Tend$ ,  $TR$ ,  $SME$ ,  $Torque$ ,  $TR$ ,  $BD$ ,  $PH$ ,  $WAI$ , and  $WSI$ ) were determined  
306 (Table 4). Reduction of quantified mycotoxins during the extrusion process is expressed as a  
307 percentage reduction concerning its initial concentration in the whole grain triticale flour, and



308 it is used in all the performed statistical analyses. Process variables (*SS*, *FR*, and *MC*) were  
309 varied according to BB experimental design (Table 4) and the range of observed responses  
310 was: *P* from 2.7 to 7.9 MPa, *Tend* from 113 to 151 °C, *SME* from 66.9 to 125 kWh/t, *Torque*  
311 from 39.6 to 59.4 Nm, mean retention time in the barrel (*TR*) from 4 to 11 s, *BD* from 0.538 to  
312 0.596 g/mL, *PH* from 6.7 to 19.4 kg, *WAI* from 3.5 to 5.4 g/g and *WSI* from 9.1 to 12.5 g/100  
313 g. Reduction of all investigated mycotoxins was achieved in all trials (extruded product  
314 samples) (Table 4). Reduction of DON ranged from 0.12 to 16.6%, while for 3-AcDON, 15-  
315 AcDON and HT-2, ranged from 1.7 to 32.8%, from 1.7 to 45.7%, and from 24.3 to 60.5%,  
316 respectively. Further, the reduction of TEN ranged from 1.7 to 21.2%, while for AME ranged  
317 from 53.2 to 91.8%. The maximum reduction rates (TS-15) for DON and AME of 16.6 and  
318 91.8%, respectively, were obtained at the following process parameters: *SS*=500 rpm, *FR*= 26  
319 kg/h, and *MC*=20 g/100 g. At the highest screw speed (800 rpm), the lowest feed rate (22  
320 kg/h), and a medium moisture content of the raw material (25 g/100 g), the highest reduction  
321 rate (32.8%) of 3-AcDON was achieved (TS-6), while the maximum reduction rate of 45.7%  
322 for 15-AcDON (TS-1) was obtained at the highest screw speed (800 rpm), the medium feed  
323 rate (26 kg/h), and at the highest moisture content of the raw material (30 g/100 g). Further, at  
324 the medium screw speed (650 rpm), the highest feed rate (30 kg/h) and the lowest moisture  
325 content of whole grain triticale flour (20 g/100 g), the highest reduction rate (60.5%) of HT-2  
326 toxin was obtained (TS-14). Regarding TEN, its maximum reduction rate (TS-12) during the  
327 extrusion process of 21.2% was achieved at the highest screw speed (800 rpm), the medium  
328 feed rate (26 kg/h), and at the lowest moisture content (20 g/100 g) of the raw material. If  
329 several mycotoxins are found in the raw material, the substantial aim is to minimize their  
330 concentrations to the lowest possible level, with a modest effect on the quality of the final  
331 product. Having this in mind, the implementation of a suitable mathematical methodology is  
332 vital for optimizing the quality of the final result.

333

334 

### 3.4 Principal component analysis

335 Firstly, the PCA analysis pursued to the acquired experimental data set has illustrated a  
336 partitioning among samples, as suggested by the factor variables and it was applied as a tool  
337 in exploratory data analysis to describe and distinguish response variables (Fig. 2). The  
338 conclusion of the PCA analysis interpreted the first three principal components, counting for  
339 67.3% of the total variance, which can be perceived as sufficient for data explanation. *Tend*,  
340 *P*, *SME*, Torque, *WAI*, reduction of TEN (*rTEN*) and *rAME* had been more potent for the  
341 primary principal component evaluation (contributing: 15.8; 11.0; 14.0; 9.3; 17.0; 8.3 and  
342 8.9%, accordingly, based on correlations), while *P*, *BD*, *WSI*, *r3-AcDON*, *r15-AcDON*, *rHT-2*  
343 and *rTEN* had been more crucial for the second principal component computation (9.9; 15.5;  
344 14.8; 23.3; 12.3; 10.9 and 7.7%, respectively). The most powerful factors for PC3 calculation  
345 were Torque, *PH*, *rDON* and *r3-AcDON*, and *rHT-2* (with a share of 10.3; 24.3; 26.4; 10.5  
346 and 8.1%, individually). The PCA plot (Fig. 1) pointed out well segregation between samples.  
347 Samples acquired by higher screw speed are positioned at the right side of the chart; these  
348 samples are classified by higher *P*, *RT*, *Tend*, Torque, *SME*, and *WAI*, and also by the  
349 augmented reduction of DON, AME, and TEN content.

350

351 

### 3.5 Response surface method

352 ANOVA evaluation was performed on the developed SOP models, and each of them was  
353 investigated on the effects of input variables (Table 5). The analysis demonstrated that the  
354 linear terms of *SS* and *MC* were the most significant variables in the SOP model for *Tend*  
355 computation (statistically significant at  $p<0.01$  and  $p<0.05$ , accordingly) while the impact of  
356 interchange term  $SS \times MC$  was significant at  $p<0.05$  level. *P* evaluation was mostly affected  
357 by the linear terms of *SS* and *FR* in the SOP model (statistically significant at  $p<0.01$  and

358  $p < 0.05$  levels, respectively). The linear terms of  $SS$  and  $FR$ , as well as the quadratic term of  
359  $FR$  were the most influential for  $SME$  calculation (statistically significant at  $p < 0.01$  level),  
360 while the linear term of  $MC$  and the interactive terms  $SS \times MC$  and  $MC \times FR$  were influential  
361 at the statistically significant level of  $p < 0.05$ .  $Torque$  was mostly impacted by the linear terms  
362 of  $SS$  and  $MC$  in the SOP model (statistically significant at  $p < 0.01$  level), while the linear  
363 term of  $FR$  and the quadratic terms of  $SS$  and  $FR$  significantly contributed to  $Torque$   
364 evaluation ( $p < 0.05$ ).  $BD$  and  $PH$  were altered by the non-linear term of  $SS \times MC$  ( $p < 0.05$   
365 level), while  $PH$  was also influenced by the linear term of  $MC$  (statistically significant at  
366  $p < 0.01$  level).

367 The prior studies were focused on the increase of  $SS$ , which conducted the lowered  $BD$  of  
368 extrudates (Ding et al., 2006; Fili et al., 2012; Gulati et al., 2016). This was also displayed in  
369 Table 5. The higher  $SS$  augmented the elasticity of the dough in the extruder tube, which  
370 decrease  $BD$  (Fletcher et al., 1985). The raise of  $MC$  guided to an increase in  $BD$  (Ding et al.  
371 2005; Gulati et al., 2016; Liu et al., 2011), and this was also corroborated in this study (Table  
372 5).

373  $WAI$  and  $WSI$  were impacted by the quadratic term of  $SS$ , while  $WAI$  was also influenced  
374 by the linear term of  $SS$  (statistically significant at  $p < 0.01$  level). These results are in consent  
375 with earlier studies for the following samples: an extruded mixture of maize bite and spell  
376 (Jozinović et al., 2016), quinoa (Dogan and Karwe, 2003), amaranthus (Menegassi et al.,  
377 2011), sorghum (Mahasukhonthachat et al., 2010), corn grits with buckwheat and chestnut  
378 (Jozinović, 2012), and corn-wheat extrudate (Sobota et al. 2010).

379 The analysis explained that the quadratic terms of  $MC$  were the most effective for  $r3$ -  
380  $AcDON$  and  $rHT-2$  calculation in SOP models (statistically significant at  $p < 0.05$  level).  $rTEN$   
381 was mostly influenced by the linear terms of  $SS$  and  $MC$ , and also by the quadratic term of  $SS$   
382 and the non-linear term of  $SS \times FR$  in the SOP model ( $p < 0.01$  level), while the linear term of

383 *FR* and the non-linear term of quadratic term of *SS* × *MC* significantly contributed to *rTEN*  
 384 calculation, statistically significant at  $p < 0.05$  level. *rAME* was mostly influenced by the linear  
 385 term of *SS* in SOP model computation ( $p < 0.05$ ).

386 All SOP models had an insignificant lack of fit tests, which means that all the models  
 387 represented the data satisfactorily. The  $r^2$  values were very suitable and showed a good fit of  
 388 the model to experimental results.

389

### 390 3.6 Optimization study of the extruder parameters, performed by standard score

391 The optimal score was determined by averaging the scores for all mycotoxins reduction  
 392 variables:

$$393 \text{ Score (MC,FR,SS)} = \frac{\overline{rDON} + \overline{r3-AcDON} + \overline{r15-AcDON} + \overline{rHT-2} + \overline{rTEN} + \overline{rAME}}{6} \quad (4)$$

394 The maximum score function displayed the optimal factor variables, and also the optimum  
 395 for mycotoxins reduction variables. Standard score evaluation results were presented in Fig. 3.  
 396 The best scores were reached in sample TS-14, while the optimized parameters were as  
 397 follows: *SS* = 650 rpm, *FR* = 30 kg/h, and *MC* = 20 g/100 g. The obtained parameters for  
 398 extrusion process were: *Tend* = 151 °C, *P* = 5.9 MPa, *SME* = 107.8 Wh/kg, *Torque* = 59.4 Nm  
 399 and *RT* = 7.5 s, while the physico-chemical properties of the optimal sample were: *BD* = 0.589  
 400 g/mL, *PH* = 19.4 kg, *WAI* = 5.0 g/g and *WSI* = 9.1 g/100 g (Table 4). The reduction rates of  
 401 examined mycotoxins at optimal extrusion conditions were as follows: 9.5, 27.8, 28.4, 60.5,  
 402 12.3, and 85.7%, for DON, 3-AcDON, 15-AcDON, HT-2, TEN, and AME, respectively. The  
 403 other two samples which were close to optimal score were samples TS-13 and TS-2, which  
 404 gained scores of 0.689 and 0.657, respectively (Fig. 3). Sample TS-13 was produced using  
 405 extruder parameters: *SS* = 650 rpm, *FR* = 22 kg/h and *MC* = 20 g/100 g, while sample TS-2  
 406 was obtained using parameters: *SS* = 650 rpm, *FR* = 22 kg/h and *MC* = 30 g/100 g. The  
 407 physical-chemical properties of sample TS-13 are characterized by the lowest pellet hardness

408 of 6.7 kg compared to other produced pellets, while the values for *WAI* and *WSI* were  
409 approximately at the same level as in the sample TS-14. Sample TS-2 had the largest bulk  
410 density (g/mL) relative to other extruded produced and medium hardness, while *WAI* was one  
411 unit lower than *WAI* of sample TS-14 (Table 4). Regarding mycotoxins reduction by above  
412 mentioned extrusion conditions, in extrudates TS-13, reduction by 14.9, 30.8, 35.5, 45.7, 12.9,  
413 and 81.4 %, for DON, 3-AcDON, 15-AcDON, HT-2, TEN, and AME, respectively were  
414 achieved, while in samples TS-2 reduction of 13.0, 32.4, 5.5, 57.0, 12.4 and 83.4% for DON,  
415 3-AcDON, 15-AcDON, HT-2, TEN and AME, respectively were obtained.

416 As some mycotoxins are highly toxic, maximum limit levels have been established to  
417 protect consumers' health. Among examined mycotoxins, Commission regulation (EC) No.  
418 1881/2006 set maximum levels in foodstuffs (EC, 2017) just for DON. Further, Commission  
419 recommendation No. 165/2013 (EU, 2013) on the presence of T-2 and HT-2 toxin in cereals  
420 and cereal products has issued recommended levels for the sum of HT-2 and T-2, which is 50  
421  $\mu\text{g}/\text{kg}$ . Accordingly to the presence of the mycotoxins in original triticale flour before the  
422 extrusion process, only DON and HT-2 content can be used for evaluation of possible risks  
423 for human health. The maximum permitted level for DON stated in EC (2017) for cereals  
424 intended for direct human consumption, cereal flour is 750  $\mu\text{g}/\text{kg}$ . The content of DON in  
425 triticale flour before extrusion in our study was 274.4  $\mu\text{g}/\text{kg}$ , and after the process, the DON  
426 content was reduced under optimized conditions for a maximum of 16.6%. Also HT-2 content  
427 was lower as 50  $\mu\text{g}/\text{kg}$  before and after the extrusion. DON and HT-2 content in triticale flour  
428 can be evaluated as non-risk flour for human consumption. Evaluation of the other  
429 mycotoxins content cannot be done, since maximum permitted level or indicative recommend  
430 level data for them are not known.

431 The obtained results in this study may have a great contribution to the further selection of  
432 appropriate extrusion parameters depending on the mycotoxins present in whole grain triticale

433 flour. It is well known that all of the investigated mycotoxins (DON, 3-AcDON, 15-AcDON,  
434 HT-2, AME, and TEN) may potentially affect human and animal health. Most of the data on  
435 their toxicity concern their effects when present alone. The investigated trichothecenes (DON,  
436 3-AcDON, 15-AcDON, and HT-2) are potent inhibitors of protein, DNA, and RNA synthesis,  
437 causing teratogenic, neurotoxic, embryotoxic, and immunosuppressive effects. The effects of  
438 short-term consumption of contaminated food could lead to nausea and vomiting followed by  
439 abdominal pain, diarrhea, headache, dizziness, and fever (He et al., 2007; Chen et al., 2020).  
440 From the perspective of human health, only DON is classified by International Agency for  
441 Research on Cancer in Group 3 not classifiable as to its carcinogenicity (Ostry, Mlir, Toman,  
442 & Grosse, 2017). *Alternaria* toxins may have acute or chronic toxic effects and pronounced  
443 fetotoxic, teratogenic, and mutagenic effects (Bhunja, 2018). Hence, in terms of human and  
444 animal health protection, any achieved reduction of mycotoxins contamination, both  
445 individual and combined mycotoxins, is of great importance. If whole grain triticale flour is  
446 contaminated with a high level of the following individual mycotoxins DON, 3-AcDON, 15-  
447 AcDON, and HT-2, for their maximum reduction, extrusion processing parameters used for  
448 the production of samples TS-15, TS-6, TS-1, and TS-14 (Table 4), should be applied,  
449 respectively. Further, if a sample is contaminated with *Alternaria* toxins, extrusion processing  
450 parameters for the production of TS-12, should be used for the maximum reduction of AME  
451 and TEN. Contamination of whole triticale flour with more than one mycotoxin would require  
452 a compromise solution such as the choice of extrusion parameters that most reduce the  
453 concentrations of mycotoxins present and at the same time achieve a satisfactory quality of  
454 the final products. In the present study, extrusion processing parameters applied for the  
455 sample TS-14, followed by TS-13 and TS-6, resulted in the best reduction of the sum of  
456 mycotoxins present in whole grain triticale flour.

457

### 458 3.7. Comparison of the obtained results with the literature data

459 The obtained results in this study could not be completely compared to the published data,  
460 since to the best of the authors' knowledge there is no previously published data regarding the  
461 fate of a larger number of co-occurred mycotoxins in triticale during the extrusion process.  
462 Regarding the reduction of *Fusarium* toxins content by the extrusion process, the so far  
463 published data relate mainly to the reduction of DON in wheat and FBs in maize  
464 (Schaarschmidt & Fauhl-Hassek, 2018; 2021). Changes in DON content during the extrusion  
465 process (twin-screw extruder) of whole grain wheat flour ranged from +1 to – 23%, while the  
466 reduction rate of DON during the extrusion process (twin-screw extruder) of soaked wheat  
467 grains ranged from 6 to 10% (Schaarschmidt & Fauhl-Hassek, 2018). Reduction of DON  
468 content during the extrusion process of whole grain triticale flour (0.12 – 16.6%) in this study  
469 is in agreement with the published findings so far. On the other hand, by extrusion of spiked  
470 wheat grits with laboratory-scale single-screw extruder, reduction of DON content was ranged  
471 from 3 to 60%, depending on applied extrusion process parameters (Wu, Lohrey, Cramer,  
472 Yuan, & Humpf, 2011). Further, Pleadin et al. (2019) reported, that depending on applied  
473 temperature profiles in dosing/compression/ejection zone of the laboratory scale single-screw  
474 extruder (135/150/150 °C; 135/170/170 °C and 135/190/190 °C), the following reduction rates  
475 of DON were obtained: 51, 61 and 71 % for wheat; 73, 80 and 87% for oat; and 55, 60 and  
476 66% for maize. Similar results were obtained regarding DON reduction during the extrusion  
477 process of maize (Schaarschmidt & Fauhl-Hassek, 2021). Namely, by extrusion of maize grits  
478 using laboratory-scale twin-screw extruder, the reduction rate of DON was ranged from 22 to  
479 53%, while by using laboratory-scale single-screw extruder for extrusion of whole grain maize  
480 grits, its reduction was from 55 to 66%. Contrary to the above mentioned findings, by  
481 extrusion of maize grits with a pilot-scale twin-screw extruder, the reduction rates of DON  
482 were ranged from 3 to 13%. The so far published study regarding the fate of DON during the

483 extrusion process indicated, that the reduction rate of DON increased at higher temperatures,  
484 and lower moisture content of the raw material. Our findings are in agreement with the  
485 available data on the fate of DON during extrusion, since the maximum reduction rate of  
486 16.6% (T-15) was achieved at a similar condition (Table 4).

487 Concerning the fate of *Alternaria* toxins during the extrusion process, only the fate of AME  
488 can be compared to our previous study (Janić Hajnal et al., 2016). Namely, the reduction rates  
489 of AME depending on applied process parameters in this study ranged between 53.2 to 91.8%,  
490 while in our previous study its reduction rate was very similar (62.8 to 94.5%). Moreover, the  
491 maximum reduction rate of AME of 91.8% in the present study (pilot-scale twin-screw  
492 extruder) is very similar to the obtained reduction rate of AME (94.5%) by extrusion of whole  
493 grain wheat flour using a pilot-scale single screw extruder (Janić Hajnal et al., 2016). In the  
494 present study, AME reduction was mostly affected by the linear term of screw speed of twin-  
495 screw extruder ( $p < 0.05$ ), while in our previous study the level of AME reduction was mostly  
496 influenced by the linear term of moisture content of the whole grain wheat flour and the  
497 quadratic term of screw speed of single-screw extruder (Janić Hajnal et al., 2016). Findings of  
498 present and previous studies indicate that the level of reduction of AME content during the  
499 extrusion of small grain cereals (whole grain triticale flour, whole grain wheat flour) is very  
500 similar by using both types of extruders, although the process parameters, to achieve the  
501 maximum reduction of the AME content differ among the types of extruders used.

502

#### 503 **4. Conclusions**

504 To the best of our knowledge, the results of this study represent the first report regarding  
505 the fate of a larger number of mycotoxins, as well as the first data about the fate of some less  
506 studied mycotoxins (3-AcDON, 15-AcDON, HT-2, TEN, and AME) during the extraction  
507 process of whole grain triticale flour. The best standard scores were obtained by using



508 extrusion process parameters with the medium screw speed , the highest feed rate , and the  
509 lowest moisture content of raw material , which provide the optimal reduction rates of present  
510 mycotoxins in the final product.. In brief, due to the combined action of heat, pressure, and  
511 shear force, the extrusion process has conditionally a high potential for mycotoxins reduction.  
512 However, based on the so far published data, as well as on the obtained results, it can be  
513 concluded that due to the complex interaction of the various parameters, the effect of the  
514 extrusion process on different mycotoxins still needs to be determined in detail for each  
515 combination of ingredient composition, as well as for applied parameters setting. In the case  
516 of contamination of the raw material with a large number of mycotoxins, it is not possible to  
517 achieve the maximum reduction of the content of each present mycotoxins by extrusion  
518 process. For this reason, there is a need to find a compromise solution, i.e. optimal extrusion  
519 conditions that will provide a satisfactory reduction of mycotoxins present, as well as a  
520 satisfactory quality of the final products.

521

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528

## 529 **Declaration of interests**

530 The authors declare that they have no known competing financial interests or personal  
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532

533 **References**

- 534 Agriopoulou, S., Stamatelopoulou, E., & Varzakas, T. (2020). Advances in Occurrence,  
535 Importance, and Mycotoxin Control Strategies: Prevention and Detoxification in Foods.  
536 *Foods*, 9(2), 137. <https://doi.org/10.3390/foods9020137>.
- 537 Anderson, R. A., Conway, H., & Peplinski, A. J. (1970). Gelatinization of corn grits by roll  
538 cooking, extrusion cooking and steaming. *Starch-Stärke*, 22, 130–135.  
539 <https://doi.org/10.1002/star.19700220408>.
- 540 Babič, J., Tavčar-Kalcher, G., Celar, F. A., Kos, K., Knific, T., & Jakovac-Strajn, B. (2021).  
541 Occurrence of *Alternaria* and Other Toxins in Cereal Grains Intended for Animal Feeding  
542 Collected in Slovenia: A Three-Year Study. *Toxins*, 13(5), 304.  
543 <https://doi.org/10.3390/toxins13050304>.
- 544 Bhunia, A. K. (2018). Molds and mycotoxins. In *Foodborne Microbial Pathogens* (pp. 167–  
545 174). Springer, New York, NY.
- 546 Bullerman, L. B., & Bianchini, A. (2007). Stability of mycotoxins during food processing.  
547 *International journal of food microbiology*, 119(1-2), 140-146.  
548 <https://doi.org/10.1016/j.ijfoodmicro.2007.07.035>.
- 549 Chen, P., Xiang, B., Shi, H., Yu, P., Song, Y., & Li, S. (2020). Recent advances on type A  
550 trichothecenes in food and feed: Analysis, prevalence, toxicity, and decontamination  
551 techniques. *Food Control*, 118, 107371. <https://doi.org/10.1016/j.foodcont.2020.107371>.
- 552 Clark, P. M., Behnke, K. C., & Poole, D. R. (2007). Effects of marker selection and mix time  
553 on the coefficient of variation (mix uniformity) of broiler feed. *The Journal of Applied*  
554 *Poultry Research*, 16(3), 464–470. <https://doi.org/10.1093/japr/16.3.464>.
- 555 Ding Q.B., Ainsworth P., Tucker G., & Marson H. (2005). The effect of extrusion conditions  
556 on the physicochemical properties and sensory characteristics of rice-based expanded

- 557 snacks. *Journal of Food Engineering*, 66(3), 283–289.  
558 <https://doi.org/10.1016/j.jfoodeng.2004.03.019>.
- 559 Ding Q.B., Ainsworth P., Plunkett A., Tucker G., & Marson H. (2006). The effect of  
560 extrusion conditions on the functional and physical properties of wheat-based expanded  
561 snacks. *Journal of Food Engineering*, 73(2), 142–148.  
562 <https://doi.org/10.1016/j.jfoodeng.2005.01.013>.
- 563 Dogan H., & Karwe M. (2003). Physicochemical properties of quinoa extrudates. *Food*  
564 *Science and Technology International*, 9(2), 101–114.  
565 <https://doi.org/10.1177/1082013203009002006>.
- 566 EC. (2017). Commission Regulation (EU) No 1881/2006 of 19 December 2006 setting  
567 maximum levels for certain contaminants in foodstuffs. Consolidated version from  
568 28.07.2017 including amendments. Retrieved from [http://eur-lex.europa.eu/legal-](http://eur-lex.europa.eu/legal-content/EN/TXT/?uri=CELEX:02006R1881-20170728)  
569 [content/EN/TXT/?uri=CELEX:02006R1881-20170728](http://eur-lex.europa.eu/legal-content/EN/TXT/?uri=CELEX:02006R1881-20170728).
- 570 EFSA (2016) European Food Safety Authority. Scientific report on the dietary exposure  
571 assessment to *Alternaria* toxins in the European population. *EFSA Journal* 14(12):4654, 1-  
572 32. doi:10.2903/j.efsa.2016.4654
- 573
- 574 EU. (2013). Commission Recommendation of 27 March 2013 on the presence of T-2 and HT-  
575 2 toxin in cereals and cereal products (2013/165/EU). *Official Journal*, L91, 12–14.  
576 Retrieved from [http://eur-lex.europa.eu/legal-content/EN/TXT/PDF/?uri=CELEX:](http://eur-lex.europa.eu/legal-content/EN/TXT/PDF/?uri=CELEX:32013H0165)  
577 [32013H0165](http://eur-lex.europa.eu/legal-content/EN/TXT/PDF/?uri=CELEX:32013H0165).
- 578 FAO (2019) Food and Agriculture Organization of the United Nations. Retrieved from  
579 <http://www.fao.org/faostat/en/#data>. Accessed March 31, 2021

- 580 Filli K.B., Nkama I., Jideani V.A., & Abubakar U.M. (2012). The Effect of Extrusion  
581 Conditions on the Physicochemical Properties and Sensory Characteristics of Millet –  
582 Cowpea Based Fura. *European Journal of Nutrition & Food Safet*, 2(1), 1–23.  
583 <https://www.journalejnfs.com/index.php/EJNFS/article/view/30012>.
- 584 Fletcher S.I., Richmond P., & Smith A.C. (1985). An experimental study of twin-screw  
585 extrusion cooking of maize grits. *Journal of Food Engineering*, 4(4), 291–312.  
586 [https://doi.org/10.1016/0260-8774\(85\)90009-3](https://doi.org/10.1016/0260-8774(85)90009-3).
- 587 Gagi, V. (2018). Triticale crop and contamination with mycotoxins under the influence of  
588 climate change—Global study. *Journal of Hygienic Engineering and Design*, 23, 30-45,  
589 UDC 633.1-152.75:632.4]:551.583(498).
- 590 Gulati P., Weier, S. A., Santra, D., Subbiah J., & Rose D. (2016). Effects of feed moisture and  
591 extruder screw speed and temperature on physical characteristics and antioxidant activity  
592 of extruded proso millet (*Panicum miliaceum*) flour. *International Journal of Food Science  
593 and Technology*, 51(1), 114–122. <https://doi.org/10.1111/ijfs.12974>.
- 594 He, J., Yang, R., Zhou, T., Tsao, R., Young, J. C., Zhu, H., Li, X-Z., & Boland, G. J. (2007).  
595 Purification of deoxynivalenol from *Fusarium graminearum* rice culture and mouldy corn  
596 by high-speed counter-current chromatography. *Journal of Chromatography A*, 1151(1-2),  
597 187-192. <https://doi.org/10.1016/j.chroma.2007.01.112>.
- 598 Hojnik, N., Modic, M., Tavčar-Kalcher, G., Babič, J., Walsh, J.L., & Cvelbar, U. (2019).  
599 Mycotoxin Decontamination Efficacy of Atmospheric Pressure Air Plasma. *Toxins*, 11(4),  
600 219. <https://doi.org/10.3390/toxins11040219>.
- 601 ISO (2009). International Organization for Standardization. ISO 712/2009: Cereals and cereal  
602 products—Determination of moisture content (Reference method). ISO, Geneva,  
603 Switzerland.

- 604 Janić Hajnal, E., Orčić, D., Torbica, A., Kos, J., Mastilović, J., & Škrinjar, M. (2015).  
605 *Alternaria* toxins in wheat from the Autonomous Province of Vojvodina, Serbia: a  
606 preliminary survey. *Food Additives & Contaminants: Part A*, 32(3), 361–370.  
607 <https://doi.org/10.1080/19440049.2015.1007533>.
- 608 Janić Hajnal, E., Čolović, R., Pezo, L., Orčić, D., Vukmirović, Đ., & Mastilović, J. (2016).  
609 Possibility of *Alternaria* toxins reduction by extrusion processing of whole wheat flour.  
610 *Food chemistry*, 213, 784–790. <https://doi.org/10.1016/j.foodchem.2016.07.019>-
- 611 Janić Hajnal, E., Mastilović, J., Bagi, F., Orčić, D., Budakov, D., Kos, J., & Savić, Z. (2019).  
612 Effect of wheat milling process on the distribution of *Alternaria* toxins. *Toxins*, 11(3),  
613 139. <https://doi.org/10.3390/toxins11030139>.
- 614 Jozinović, A., Šubarić, D., Ačkar, Đ., Babić, J., Klarić, I., Kopjar, M., & Valek Lendić, K.  
615 (2012). Influence of buckwheat and chestnut flour addition on properties of corn  
616 extrudates. *Croatian Journal of Food Science and Technology*, 4, 26–33.  
617 <https://hrcak.srce.hr/84717>.
- 618 Jozinović, A., Šubarić, D., Ačkar, Đ., Babić, J., & Miličević, B. (2016). Influence of spelt  
619 flour addition on properties of extruded products based on corn grits. *Journal of Food*  
620 *Engineering*, 172, 31–37. <https://doi.org/10.1016/j.jfoodeng.2015.04.012>.
- 621 Kojić, J. S., Ilić, N. M., Kojić, P. S., Pezo, L. L., Banjac, V. V., Krulj, J. A., & Bodroža  
622 Solarov, M. I. (2019). Multiobjective process optimization for betaine enriched spelt flour  
623 based extrudates. *Journal of Food Process Engineering*, 42(1), Article e12942.  
624 <https://doi.org/10.1111/jfpe.12942>. Liu, C., Zhang, Y., Liu, W., Wan, J., Wang, W., Wu, L.,  
625 Zuo, N., Zhou, Y. & Yin, Z. (2011). Preparation, physicochemical and texture properties of  
626 texturized rice produce by improved extrusion cooking technology. *Journal of Cereal*  
627 *Science*, 54, 473-480. <https://doi.org/10.1016/j.jcs.2011.09.001>.

- 628 Liu, Y., Yamdeu, J. H. G., Gong, Y. Y., & Orfila, C. A. (2020). Review of postharvest  
629 approaches to reduce fungal and mycotoxin contamination of foods. *Comprehensive*  
630 *Reviews in Food Science and Food Safety*, 19(4), 1521–1560.  
631 <https://doi.org/10.1111/1541-4337.12562>.
- 632 Mahasukhonthachat K., Sopade P., & Gidley M. (2010). Kinetics of starch digestion and  
633 functional properties of twin-screw extruded sorghum. *Journal of Cereal Science*, 51(3),  
634 392–401. <https://doi.org/10.1016/j.jcs.2010.02.008>.
- 635 Massarolo, K. C., Mendoza, J. R., Verma, T., Kupski, L., Eliana Badiale-Furlong, E., &  
636 Bianchini, A. (2021). Fate of aflatoxins in cornmeal during single-screw extrusion: A  
637 bioaccessibility approach. *LWT-Food Science and Technology*, 138, 110734.  
638 <https://doi.org/10.1016/j.lwt.2020.110734>.
- 639 Menegassi, B., Pilosof, A.M., & Areas, J.A.G. (2011). Comparison of properties of native and  
640 extruded amaranth (*Amaranthus cruentus* L.–BRS Alegria) flour. *LWT-Food Science and*  
641 *Technology*, 44(9), 1915–1921. <https://doi.org/10.1016/j.lwt.2011.04.008>.
- 642 Oliveira, P. M., Zannini, E., & Arendt, E. K. (2014). Cereal fungal infection, mycotoxins, and  
643 lactic acid bacteria mediated bioprotection: from crop farming to cereal products. *Food*  
644 *microbiology*, 37, 78–95. <https://doi.org/10.1016/j.fm.2013.06.003>.
- 645 Ostry, V., Mlir, F., Toman, J., & Grosse, Y. (2017). Mycotoxins as human carcinogens – the  
646 IARC Monograph classification. *Mycotoxin Research*, 33, 65–73.  
647 <https://doi.org/10.1007/s12550-016-0265-7>.
- 648 Pleadin, J., Babić, J., Vulić, A., Kudumija, N., Aladić, K., Kiš, M., Jaki Tkalec, V.,  
649 Škrivanko, M., Lolić, M., & Šubarić, D. (2019). The effect of thermal processing on the  
650 reduction of deoxynivalenol and zearalenone cereal content. *Croatian Journal of Food*  
651 *Science and Technology*, 11(1), 44–51. <https://doi.org/10.17508/CJFST.2019.11.1.06>.

- 652 Schaarschmidt, S., & Fahl-Hassek, C. (2018). The fate of mycotoxins during the processing  
653 of wheat for human consumption. *Comprehensive Reviews in Food Science and Food*  
654 *Safety*, 17(3), 556–593. <https://doi.org/10.1111/1541-4337.12338>.
- 655 Schaarschmidt, S., & Fahl-Hassek, C. (2021). The fate of mycotoxins during secondary food  
656 processing of maize for human consumption. *Comprehensive Reviews in Food Science and*  
657 *Food Safety*, 20(1), 91–148. <https://doi.org/10.1111/1541-4337.12657>.
- 658 Singha, P., &  
659 Muthukumarappan, K. (2017). Effects of processing conditions on the system parameters  
660 during single screw extrusion of blend containing apple pomace. *Journal of Food Process*  
*Engineering*, 40(4), Article e12513. <https://doi.org/10.1111/jfpe.12513>.
- 661 Singha, P., Singh, S. K., Muthukumarappan, K., & Krishnan, P. (2018). Physicochemical and  
662 nutritional properties of extrudates from food grade distiller's dried grains, garbanzo flour,  
663 and corn grits. *Food Science Nutrition*, 6(7), 1914–1926. <https://doi.org/10.1002/fsn3.769>.
- 664 Sobota A., Sykut-Domańska E., & Rzedzicki Z. (2010). Effect of extrusion-cooking process  
665 on the chemical composition of corn-wheat extrudates, with particular emphasis on dietary  
666 fibre fractions. *Polish Journal of Food and Nutrition Science*, 60(3), 251–259.  
667 [http://agro.icm.edu.pl/agro/element/bwmeta1.element.agro-cff37405-283d-4c2d-8a70-](http://agro.icm.edu.pl/agro/element/bwmeta1.element.agro-cff37405-283d-4c2d-8a70-0e35a223134c)  
668 [0e35a223134c](http://agro.icm.edu.pl/agro/element/bwmeta1.element.agro-cff37405-283d-4c2d-8a70-0e35a223134c).
- 669 STATISTICA (2013). StatSoftStatistica 13.3® software (v.Version 13.3.) (2013): Stat-Soft  
670 Inc, USA. ([www.statsoft.com](http://www.statsoft.com)).
- 671 Streit, E., Schatzmayr, G., Tassis, P., Tzika, E., Marin, D., Taranu, I., Tabuc, C., Nicolau, A.,  
672 Aprodu, I., Puel, O., & Oswald, I.P. (2012). Current situation of mycotoxin contamination  
673 and co-occurrence in animal feed—Focus on Europe. *Toxins*, 4(10), 788–809.  
674 <https://doi.org/10.3390/toxins4100788>.
- 675 Topi, D., Tavčar-Kalcher, G., Pavšič-Vrtač, K., Babič, J., & Jakovac-Strajn, B. (2019).  
676 *Alternaria* mycotoxins in grains from Albania: Alternariol, alternariol monomethyl ether,

- 677 tenuazonic acid and tentoxin. *World Mycotoxin Journal*, 12(1), 89–99.  
678 <https://doi.org/10.3920/WMJ2018.2342>.
- 679 Topi, D., Babič, J., Pavšič-Vrtač, K., Tavčar-Kalcher, G., & Jakovac-Strajn, B. (2021).  
680 Incidence of *Fusarium* Mycotoxins in Wheat and Maize from Albania. *Molecules*, 26(1),  
681 172. <https://doi.org/10.3390/molecules26010172>.
- 682 Torbica, A., Belović, M., Popović, L., & Čakarević, J. (2021). Heat and hydrothermal  
683 treatments of non-wheat flours. *Food Chemistry*, 334, 127523.  
684 <https://doi.org/10.1016/j.foodchem.2020.127523>.
- 685 Wan, J., Bingcan, C., & Rao, J. (2020). Occurrence and preventive strategies to control  
686 mycotoxins in cereal-based food. *Comprehensive Reviews in Food Science and Food*  
687 *Safety*, 19(3), 928–953. <https://doi.org/10.1111/1541-4337.12546>.
- 688 Wu, Q., Lohrey, L., Cramer, B., Yuan, Z., & Humpf, H-U. (2011). Impact of physicochemical  
689 parameters on the decomposition of deoxynivalenol during extrusion cooking of wheat  
690 grits. *Journal of Agricultural and Food Chemistry*, 59(23), 12480–12485.  
691 <https://pubs.acs.org/doi/pdf/10.1021/jf2038604>.
- 692



693 **Figures captions**

694

695 Fig. 1. Scheme of the co-rotating twin-screw extruder

696 **Fig. 2.** PCA ordination of variables based on component correlations, presented in the first  
697 and the second factor plane.

698 Abbreviations: *MC*  $\rightarrow$  moisture content (g/ 100 g); *FR*  $\rightarrow$  feed rate (kg/h); *SS*  $\rightarrow$  screw speed  
699 (rpm); *Tend* – die temperature ( $^{\circ}$ C); *P* - pressure at the die (MPa); *SME* – torque (Wh/kg); *RT*  
700  $\rightarrow$  retention time in the barrel (s), *BD*  $\rightarrow$  bulk density (g/mL); *PH*  $\rightarrow$  pellet hardness (kg); *WAI*-  
701 water absorption index (g/g); *WSI*  $\rightarrow$  water solubility index (g/100 g); *rDON* – reduction of  
702 deoxynivalenol (%); *r3-AcDON* – reduction of 3-acetyldeoxynivalenol (%); *r15-AcDON* –  
703 reduction of 15-acetyldeoxynivalenol (%); *rHT-2*  $\rightarrow$  reduction of HT-2 toxin (%); *rTEN* –  
704 reduction of tentoxin (%); *rAME* – reduction of alternariol monomethyl ether (%).

705

706 **Fig. 3.** Standard score analysis for mycotoxins reduction by extrusion processing of whole  
707 grain triticale flour.

**Table 1**

Independent extrusion parameters and their levels.

Experimental factor	Factor's level		
	low	center	high
Screw speed (rpm)	500	650	800
Feed rate (kg/h)	22	26	30
Moisture content (%)	20	25	30

**Table 2**

Recovery data of the employed analytical method based on the solvent ( $R_A$ ) and matrix-matched ( $R_E$ ) calibration curves and matrix effect (SSE).

Analytes	Spiking level ( $\mu\text{g}/\text{kg}$ ) <sup>a</sup>	Whole grain triticale flour			Extruded product		
		$R_A$ <sup>b</sup>	$R_E$ <sup>c</sup>	SSE <sup>d</sup>	$R_A$ <sup>b</sup>	$R_E$ <sup>c</sup>	SSE <sup>d</sup>
DON	50 – 400	80.3	102.3	73.3	90.2	100.1	90.2
3-AcDON	3.0 – 24	88.7	109.5	81.1	89.4	97.0	92.2
15-AcDON	3.0 – 24	92.6	95.4	97.1	99.0	101.7	97.4
HT-2	3.0 – 24	78.9	89.4	88.2	98.8	94.0	105.2
TEN	12.5 – 100	88.3	105.1	84.1	92.0	101.0	91.1
AME	12.5 – 100	32.3	103.3	31.3	31.6	98.6	32.1

<sup>a</sup>Concentration range of analytes for standard, matrix-matched calibration curves and calibration curves of spiked samples ( $\mu\text{g}/\text{kg}$ ).

<sup>b</sup> $R_A$  - Apparent recovery (%) calculated by the slope of spiked sample-prepared curve/slope of the solvent calibration curve.

<sup>c</sup> $R_E$  - Sample preparation recovery (%) calculated by the slope of spiked sample-prepared curve/slope of matrix-matched calibration curve.

<sup>d</sup>SSE-matrix effect (%) calculated by the slope of matrix-matched calibration curve/slope of the solvent calibration curve.

**Table 3**

Precision data of the examined mycotoxins.

Analytes	Spiking level ( $\mu\text{g}/\text{kg}$ )	Whole grain triticale flour		Extruded product	
		Repeatability <sup>a</sup> RSD (%)	Within-laboratory reproducibility <sup>b</sup> RSDs (%)	Repeatability <sup>a</sup> RSD (%)	Within-laboratory reproducibility <sup>b</sup> RSDs (%)
DON	50	14.6	17.8	11.9	14.1
	100	10.9	12.0	3.68	6.74
	200	10.3	11.8	2.90	4.64
	400	4.32	7.81	1.97	2.57
3-AcDON	3	11.1	15.3	8.66	10.0
	6	10.2	11.2	7.79	9.55
	12	9.38	10.3	7.85	8.74
	24	5.89	8.25	5.98	6.79
15-AcDON	3	7.71	12.7	4.04	10.9
	6	4.35	6.95	3.79	5.37
	12	3.79	6.36	3.14	4.47
	24	3.31	6.21	2.75	4.38
HT-2	3	13.8	18.0	13.55	12.8
	6	13.2	14.5	12.4	9.13
	12	7.07	7.46	5.56	8.78
	24	6.92	6.89	3.94	6.88
TEN	12.5	6.01	7.38	4.86	6.24
	25	3.27	10.1	6.26	7.35
	50	6.69	10.1	2.58	2.94
	100	3.93	4.29	3.93	5.18
AME	12.5	4.79	9.81	2.99	9.82
	25	4.86	5.53	1.92	5.68
	50	2.52	2.94	2.37	4.93
	100	4.22	9.74	4.77	9.38

<sup>a</sup>Results expressed as mean (RSD) (n = 6).<sup>b</sup>Results expressed as mean (RSD<sub>s</sub>) (n = 3×6).

**Table 4**  
Technological parameters of extrusion and reduction of mycotoxins.

Sample	Factors			Process responses					Product responses									
	<i>SS</i>	<i>FR</i>	<i>MC</i>	<i>Tend</i>	<i>P</i>	<i>SME</i>	Torque	<i>RT</i>	<i>BD</i>	<i>PH</i>	<i>WAI</i>	<i>WSI</i>	<i>rDON</i>	<i>r3-AcDON</i>	<i>r15-AcDON</i>	<i>rHT-2</i>	<i>rTEN</i>	<i>rAME</i>
TS-1	800	26	30	116	3.0	87.8	46.2	7.0	0.580	14.8	3.7	10.6	0.12	8.6	45.7	24.3	2.9	53.2
TS-2	650	22	30	118	3.1	66.9	39.6	6.0	0.596	12.7	4.1	9.2	13.0	32.4	5.5	57.0	12.4	83.4
TS-3	650	30	30	118	4.1	71.1	44.0	6.0	0.551	14.6	3.5	10.9	13.3	24.3	4.6	56.8	2.4	65.4
TS-4	500	26	30	113	2.7	68.9	44.0	6.0	0.565	13.3	3.6	11.7	13.8	31.2	37.0	44.5	9.0	61.4
TS-5	650	26	25	132	5.4	90.8	44.0	5.8	0.572	14.3	4.4	11.0	9.1	6.0	13.3	35.8	3.4	83.8
TS-6	800	22	25	132	4.4	113.5	46.2	5.0	0.571	14.7	4.5	11.0	15.1	32.8	33.6	47.0	12.6	76.3
TS-7	650	26	25	129	5.3	91.2	44.0	6.8	0.569	13.8	3.8	12.5	9.7	5.5	13.0	39.1	3.0	84.0
TS-8	800	30	25	134	4.8	94.7	55.0	5.5	0.589	14.7	4.0	11.1	8.8	30.8	35.4	47.7	4.7	76.1
TS-9	650	26	25	131	5.5	90.7	44.0	6.8	0.565	14.5	3.9	11.0	11.1	5.7	13.0	37.6	3.3	83.0
TS-10	500	22	25	129	6.1	88.6	44.0	6.0	0.547	10.5	4.1	11.1	15.4	23.6	1.7	34.9	5.2	55.5
TS-11	500	30	25	132	5.8	88.0	44.0	4.0	0.564	15.4	4.1	11.8	13.4	1.7	12.2	45.1	1.7	83.8
TS-12	800	26	20	140	5.6	125.0	57.2	6.5	0.542	13.3	5.4	11.1	15.5	18.5	16.6	26.2	21.2	88.7
TS-13	650	22	20	139	5.6	119.3	46.2	5.5	0.556	6.7	5.2	9.3	14.9	30.8	35.5	45.7	12.9	81.4
TS-14	650	30	20	151	5.9	107.8	59.4	7.5	0.589	19.4	5.0	9.1	9.5	27.8	28.4	60.5	12.3	85.7
TS-15	500	26	20	147	7.9	111.9	52.8	11	0.538	13.7	5.1	10.1	16.6	16.5	8.1	43.9	9.4	91.8

*SS*: screw speed (rpm); *FR*: feed rate (kg/h); *MC*: moisture content (g/100 g); *Tend*: die temperature (°C); *P*: pressure at the die (MPa); *SME*: specific mechanical energy(Wh/kg); Torque (Nm); *RT*:mean retention time in the barrel (s), *BD*: bulk density (g/mL); *PH*: pellet hardness (kg); *WAI*: water absorption index (g/g); *WSI*: water solubility index (g/100 g); *rDON*: reduction of deoxynivalenol (%); *r3-AcDON*: reduction of 3-acetyldeoxynivalenol (%); *r15-AcDON*: reduction of 15-acetyldeoxynivalenol (%); *rHT-2*:reduction of HT-2 toxin (%); *rTEN*: reduction of tentoxin (%); *rAME*: reduction of alternariol monomethyl ether (%).

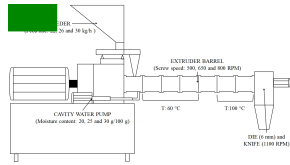
**Table 5**

ANOVA evaluation of technological parameters and reduction of mycotoxins (sum of squares).

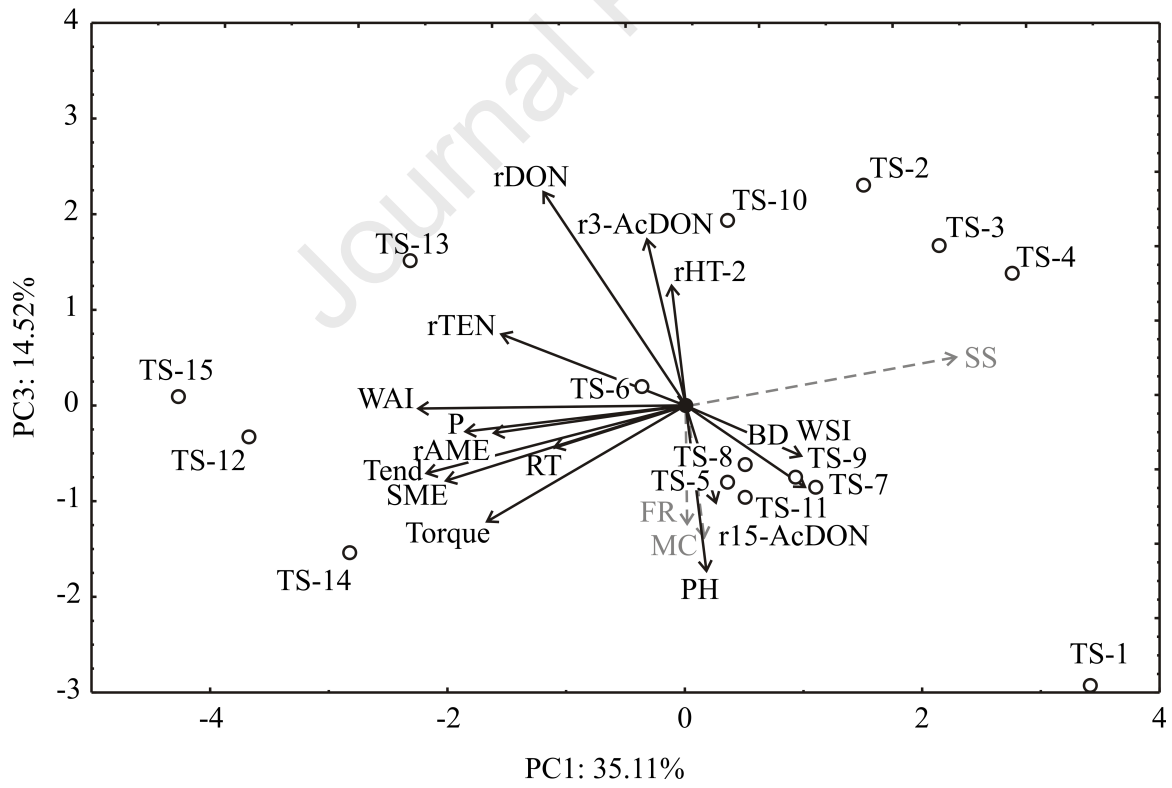
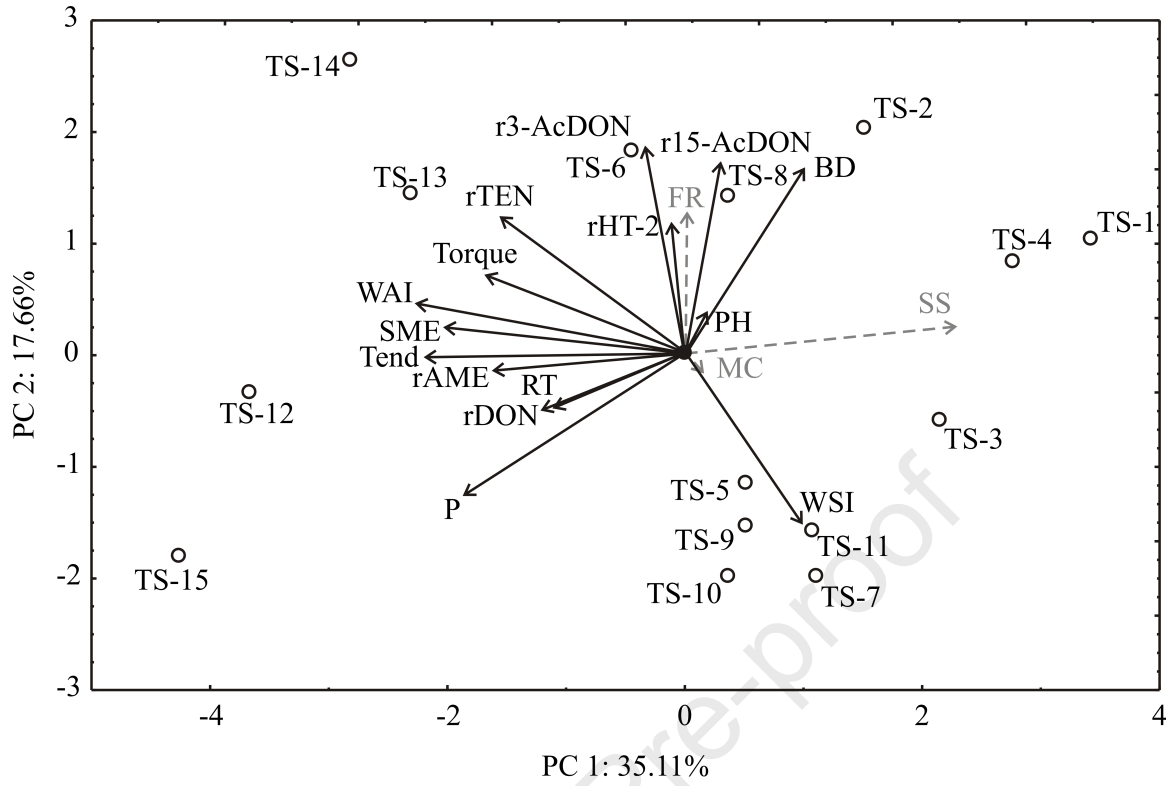
	df	<i>Tend</i>	<i>P</i>	<i>SME</i>	Torque	<i>RT</i>	<i>BD</i>	<i>PH</i>	<i>WAI</i>	<i>WSI</i>	<i>rDON</i>	<i>r3-AcDON</i>	<i>r15-AcDON</i>	<i>rHT-2</i>	<i>rTEN</i>	<i>rAME</i>
<i>SS</i>	1	1568.0 <sup>+</sup>	1869.7 <sup>+</sup>	3582.8 <sup>+</sup>	218.4 <sup>+</sup>	3.8	0.0006	0.7	4.1 <sup>+</sup>	0.9	33.4	1.1	2.2	4.9	106.4 <sup>+</sup>	886.2 <sup>*</sup>
<i>SS</i> <sup>2</sup>	1	3.4	135.1	6.1	33.8 <sup>*</sup>	4.8 <sup>**</sup>	0.0001	0.7	0.4 <sup>*</sup>	4.8 <sup>*</sup>	1.0	353.4	122.6	68.0	118.9 <sup>+</sup>	13.2
<i>MC</i>	1	36.1 <sup>*</sup>	26.3	89.1 <sup>*</sup>	87.1 <sup>+</sup>	0.0	0.0001	48.1 <sup>+</sup>	0.2 <sup>**</sup>	0.7	22.2	153.1	2.3	81.0	60.5 <sup>+</sup>	25.9
<i>MC</i> <sup>2</sup>	1	11.9	6.7	3.1	0.3	6.8 <sup>**</sup>	0.0002	0.7	0.0	2.1 <sup>**</sup>	17.8	654.0 <sup>*</sup>	0.5	644.0 <sup>*</sup>	4.3	27.7
<i>FR</i>	1	0.1	300.1 <sup>*</sup>	505.6 <sup>+</sup>	49.0 <sup>*</sup>	1.1	0.0006	2.6	0.1	0.1	48.0	39.2	653.4	68.0	32.5 <sup>*</sup>	0.4
<i>FR</i> <sup>2</sup>	1	1.9	0.1	142.5 <sup>+</sup>	33.8 <sup>*</sup>	0.0	0.0003	0.0	0.0	0.9	3.8	37.4	235.6	183.8	10.9 <sup>**</sup>	232.6
<i>SS</i> × <i>MC</i>	1	36.0 <sup>*</sup>	13.0	61.6 <sup>*</sup>	19.4 <sup>**</sup>	1.0	0.0015 <sup>*</sup>	29.4 <sup>*</sup>	0.0	0.9	8.0	6.5	9.6	55.7	21.6 <sup>*</sup>	124.3
<i>SS</i> × <i>FR</i>	1	25.0 <sup>*</sup>	175.6 <sup>**</sup>	8.4	1.2	7.6 <sup>**</sup>	0.0000	0.9	0.0	1.0	39.7	151.3	0.0	1.6	80.3 <sup>+</sup>	6.5
<i>MC</i> × <i>FR</i>	1	0.3	13.3	82.8 <sup>*</sup>	19.4 <sup>**</sup>	1.6	0.0000	5.8	0.1	0.1	4.4	99.0	18.9	22.4	4.8	203.1
Error	5	22.9	171.7	28.4	15.7	5.7	0.0009	12.8	0.3	1.7	66.0	459.9	1793.5	364.0	11.6	576.6
<i>r</i> <sup>2</sup>		0.987	0.937	0.994	0.967	0.828	0.786	0.874	0.948	0.872	0.728	0.752	0.364	0.763	0.974	0.722

*SS*: screw speed (rpm); *FR*: feed rate (kg/h); *Tend*; *MC*: moisture content (g/100 g); *Tend*: die temperature (°C); *P*: pressure at the die (MPa); *SME*: specific mechanical energy (Wh/kg); Torque (Nm); *RT*: mean retention time in the barrel (s), *BD*: bulk density (g/mL); *PH*: pellet hardness (kg); *WAI*: water absorption index (g/g); *WSI*: water solubility index (g/100 g); *rDON*: reduction of deoxynivalenol (%); *r3-AcDON*: reduction of 3-acetyldeoxynivalenol (%); *r15-AcDON*: reduction of 15-acetyldeoxynivalenol (%); *rHT-2*: reduction of HT-2 toxin (%); *rTEN*: reduction of tentoxin (%); *rAME*: reduction of alternariol monomethyl ether (%).

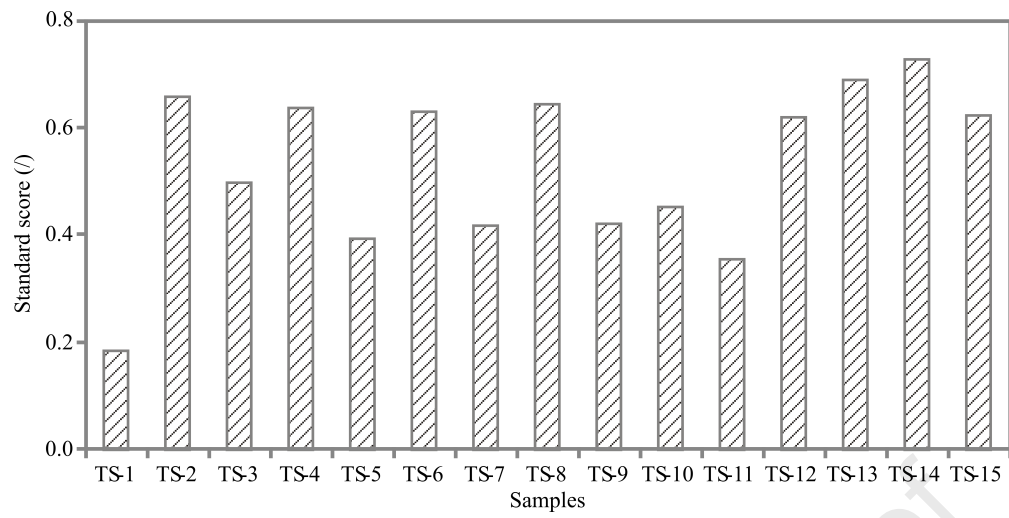
<sup>+</sup>Statistically significant at  $p < 0.01$  level; <sup>\*</sup>Statistically significant at  $p < 0.05$  level; <sup>\*\*</sup>Statistically significant at  $p < 0.10$  level.



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**Highlights**

- Reduction of mycotoxins by extrusion processing of triticale flour was investigated
- *Fusarium* and *Alternaria* toxins were analyzed by the validated LC-ESI-MS/MS method
- PCA, RSM and standard score were used to evaluate the effect of process parameters
- Optimal reduction rate of DON, 3-AcDON, 15-AcDON, HT-2, TEN and AME was determined

## **CRedit author statement**

**Elizabet Janić Hajnal:** Conceptualization, Methodology, Resources, Formal analysis, Writing - Original Draft, Writing - Review & Editing. **Janja Babič:** Formal analysis, Writing - Original Draft Writing - Review & Editing. **Lato Pezo:** Conceptualization, Writing - Original Draft, Writing - Review & Editing. **Vojislav Banjac:** Investigation, Formal analysis. **Radmilo Čolović:** Investigation, Formal analysis. **Jovana Kos:** Conceptualization, Formal analysis, Writing - Review & Editing. **Jelena Krulj:** Formal analysis. **Katarina Pavšič-Vrtač:** Formal analysis, Writing - Review & Editing. **Breda Jakovac-Strajn:** Conceptualization, Resources, Writing - Review & Editing, Supervision.

**Declaration of Competing Interest**

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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